

# Study of scavenging capacity of naringin extracted from *Citrus aurantium* peel against free radicals

## Estudio de la actividad antioxidante de la naringina extraída de la cáscara de *Citrus aurantium* contra radicales libres

Carlos E. Diaz-Urbe<sup>1</sup>, William Vallejo<sup>1</sup>, Grace Oliveros<sup>2</sup>, Amner Muñoz<sup>3</sup>

<sup>1</sup>Doctor en Ciencias, Profesor Tiempo Completo, Grupo de Investigación en Fotoquímica y Fotobiología, Universidad del Atlántico, Barranquilla-Colombia.

<sup>2</sup>Químico, Grupo de Investigación en Fotoquímica y Fotobiología, Universidad del Atlántico, Barranquilla-Colombia.

<sup>3</sup>Doctor en Ciencias, Profesor Tiempo Completo Grupo de Investigación en Química y Biología, Universidad del Norte, Barranquilla-Colombia.

E-mail: williamvallejo@mail.uniatlantico.edu.co

Recibido 10/03/2016,  
Aceptado 20/05/2016

Cite this article as: C. Diaz-Urbe, W. Vallejo, G. Oliveros, A. Muñoz, "Study of scavenging capacity of naringin extracted from *Citrus aurantium* peel against free radicals", *Prospect*, Vol 14, N° 2, 31-35, 2016.

### ABSTRACT

In this work we determined free radical scavenging of the *Citrus aurantium* peel (orange peel) extract. Naringin was isolated from the fruit peel from *Citrus aurantium* for using liquid-liquid solvent extraction method; it was characterized by spectroscopic techniques (UV-vis and FT-IR analysis).

The scavenging free radical of naringin was evaluated by monitoring radical cation bleaching of ABTS<sup>•+</sup> by visible spectroscopy at 734 nm. Assays were compared to reference antioxidants BHT,  $\alpha$ -tocopherol and Trolox. Results showed the extract from the peel of *citrus aurantium* has antioxidant activity against free radicals, under our experimental conditions, naringin extract had scavenging free radicals of 43.1 % and best reference antioxidant had scavenging free radicals of 50.4 %.

**Keywords:** Naringin; Free radicals; ABTS<sup>•+</sup>; *Citrus aurantium*.

### RESUMEN

En este trabajo determinamos la actividad antioxidante del extracto obtenido de la cáscara de *Citrus aurantium* (cáscara de naranja). El extracto de naringina fue separada de la cáscara de la fruta de la *Citrus aurantium* utilizando la extracción líquido-líquido como método de separación; el compuesto obtenido fue caracterizado por medio de espectrofotometría UV-Vis y espectroscopía infrarroja con transformada de Fourier (FT-IR).

La actividad antioxidante de la naringina fue determinada monitoreando el cambio en la concentración del catión radical ABTS<sup>•+</sup> por espectrofotometría a una longitud de onda de 734 nm; como patrones de referencia se utilizó el BHT,  $\alpha$ -tocoferol y el Trolox. Los resultados mostraron que el extracto de la cáscara de la *Citrus aurantium* presenta actividad antioxidante contra radicales; bajo nuestras condiciones experimentales, el extracto presentó un actividad antioxidante del 43.1 %, en comparación el mejor resultado para los antioxidantes de referencia fue del 50.4 %.

**Palabras clave:** Naringina; Radicales, ABTS<sup>•+</sup>; *Citrus aurantium*.

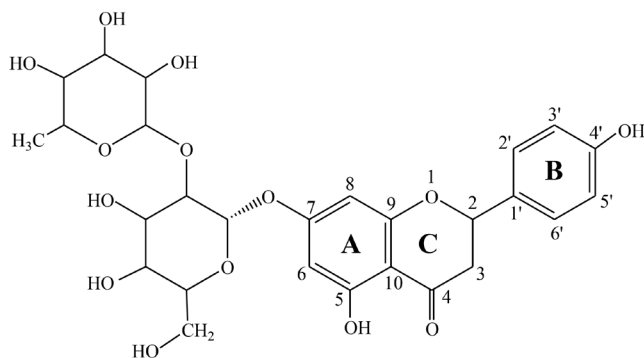
## 1. INTRODUCTION

Reactive oxygen species (ROS) have the ability to react with a large variety of biological components (e.g., lipids, sterols, proteins, DNA, and RNA) for inducing deterioration of biological systems; among ROS, hydroxyl free radical ( $\bullet\text{OH}$ ) exhibits strongest oxidative activity [1-3]. In recent years, to study the antioxidant ability of different natural compounds is an important subject of research in the field; natural compounds have possible applications in the pharmaceutical, cosmetic and food industries. Nowadays, research about  $\bullet\text{OH}$  has been focused on specific chemical species in the medicine sciences with regard to disease factors and health maintenance [4].

The antioxidant activity is the ability of a substance to inhibit the oxidative degradation of different compounds; the antioxidant capacity relies on ability of specific compounds to react to radical and non-radical species (e.g.,  $\bullet\text{OH}$ ) present in the chemical or biological environment. As a result, development of a practical assay for hydroxyl radical scavenging measurement is important, and it is a challenging task owing to the high activity, short lifetime, and low concentration of  $\bullet\text{OH}$  [5-7]. Different natural antioxidants from fruits and vegetables (e.g., carotenoids, vitamins and flavonoids) "scavenging" to  $\bullet\text{OH}$ . Among these, plant-derived flavonoids have been shown a wide range of biological activities, actually, flavonoids have demonstrated higher potential antioxidant and antibacterial activity against different ROS species, research about new natural sources of both cheap and high activity is an important research topic [8-10]. Naringin (4',5,7-trihydroxyflavanone-7- $\beta$ -L-rhamnoglucoside-(1,2)- $\alpha$ -D-glucopyranoside) is one flavanone found in the peel and juice of grapefruit and certain types of orange like *Citrus aurantium* (see figure 1); this compound has showed biological properties such as anti-inflammatory, anti-carcinogenicity and neuro-protective effect [11-13].

**Figure 1.** Chemical structure of naringin.

**Figura 1.** Estructura química de la naringina.



It is known that flavonoids from fruits extracts are scavenger to  $\bullet\text{OH}$  in solution and their possible applications as antioxidant to protect different biological systems is an important research issue, however physical-chemistry reports about the scavenger to  $\bullet\text{OH}$  that permits so study and to compare kinetical parameter to develop possible application are limited. The aim of this work was studied antioxidant capacity of the naringin obtained from inexpensive and abundant source orange peel, and besides, we tried to give relevant information about its potential natural antioxidant.

## 2. EXPERIMENTAL

### 2.1. Reagents and equipment

All reagents used in this work were analytical grade. The properties of the compound were studied by measurements of UV-Vis and Fourier transform infrared spectroscopy (FT-IR). The UV-Vis spectrum was measured in a Hewlett-Packard 8453 spectrophotometer, and the FT-IR spectra (KBr) of the compounds were recorded on a Bruker Tensor 27 spectrometer in the spectrum region between 4000 and 500  $\text{cm}^{-1}$ .

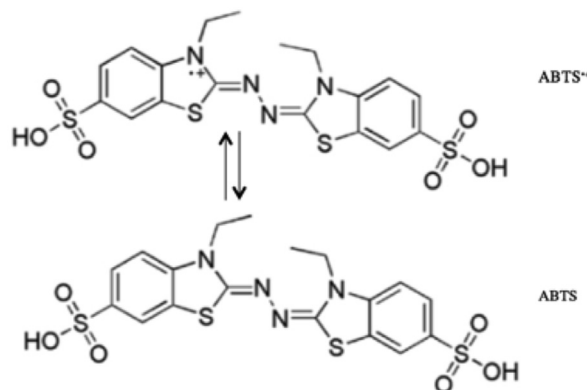
### 2.2. Isolation of naringin from *Citrus aurantium*

Sour orange (*Citrus aurantium*) fruits, in fresh and ripe state, were collected from Baranoa (latitude 10° 73' 33", longitude 74° 91' 66" – Colombia) between April and May 2013. After washing the fruits, the flavedo was mechanically separated of the albedo. Naringin was isolated from peel fruits according to the procedure described by Ikan [14]. After isolation; the solid was chemically characterized by UV-Vis and IR-FT techniques.

### 2.3 ABTS<sup>•+</sup> radical scavenging capacity

**Figure 2.** Chemical structures of ABTS and cation radical ABTS<sup>•+</sup> [16].

**Figura 2.** Estructuras químicas del ABTS y el catión radical ABTS<sup>•+</sup> [16].



The scavenging the ABTS<sup>•+</sup> radical scavenging assay was developed follow procedure described by Miller et. al. This method relies on the ability of an antioxidant to stabilize the radical cation ABTS<sup>•+</sup>, which is previously formed by the oxidation of ABTS (2,2'-azinobis (3-symphonic ethylbenzothiazoline-6 acid) and hydrogen peroxide meta-myoglobin; the results are expressed as Trolox equivalents or TEAC (Trolox Equivalent Antioxidant Capacity); figure 2 shows structures of ABTS and cation radical ABTS<sup>•+</sup> [15, 16]. Scavenging free radical of naringin was evaluated by monitoring radical cation bleaching of ABTS<sup>•+</sup> by visible spectroscopy at 734 nm; a solution 14 mg ABTS and 3.3 mg of potassium persulfate was prepared in distilled water and reacted in the dark (conditions t: 16-24 h, T: 25 °C). After that, stock solutions of the extracts using ethanol as solvent were prepared. While performing tests each 20 uL of stock solution and 2 mL of ABTS<sup>•+</sup> solution were added. The scavenging free radical (SFR %) of radical cation ABTS<sup>•+</sup> was calculated according to the following equation:

$$SFR(\%) = \left( \frac{A_o - A_f}{A_f} \right) \quad (1)$$

Where:  $A_o$  is the absorbance of uninhibited radical cation,  $A_f$  is the absorbance measured at 30 min after addition of antioxidant potential. All assays were in triplicate director.

### 3. RESULTS AND DISCUSSION

#### 3.1. Characterization of isolated naringin

The UV-vis spectrum of naringin in ethanol is shown in figure 3. Naringin exhibits two major absorption bands in the UV region, at 284 nm (band I) representing B-ring absorption (cinnamoyl system), and at 229 nm (band II), considered to be associated with the absorption involving the A ring benzoyl system, this is a typical result for flavones and flavonols. Furthermore, the FT-IR spectrum of naringin is shown in figure 4. This spectrum is characterized by vibrational bands mainly due to the OH, C-H, C=O, and C=C functional groups. The  $\nu$  (OH),  $\nu$  (C-H),  $\nu$  (C=O),  $\nu$  (C=C) for naringin appear at 3425  $\text{cm}^{-1}$ , 2957–2859  $\text{cm}^{-1}$ , 1645  $\text{cm}^{-1}$  respectively [17].

#### 3.2 ABTS<sup>•+</sup> radical scavenging capacity

The scavenging activity was evaluated by monitoring ABTS<sup>•+</sup>. Assays were compared with reference antioxidants BHT,  $\alpha$ -tocopherol and Trolox.

Figure 3. UV-Vis spectra of naringin in ethanol.  
 Figura 3. Espectro UV-Vis de la naringina.

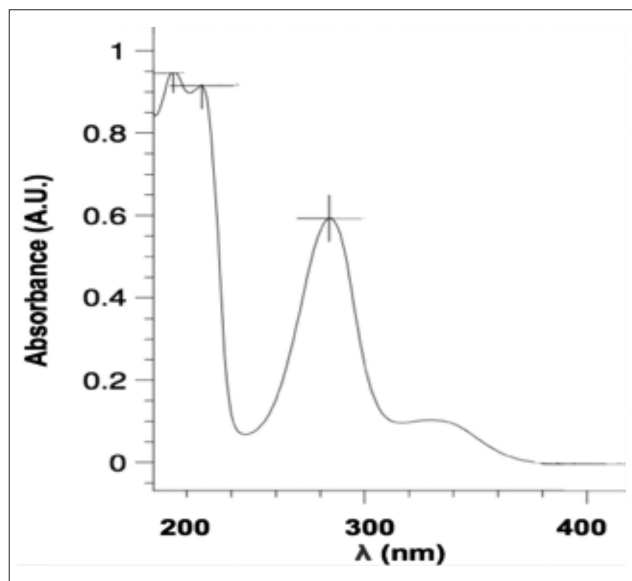


Figure 4. FT-IR spectrum of naringin.  
 Figura 4. Espectro FT-IR de la naringina.

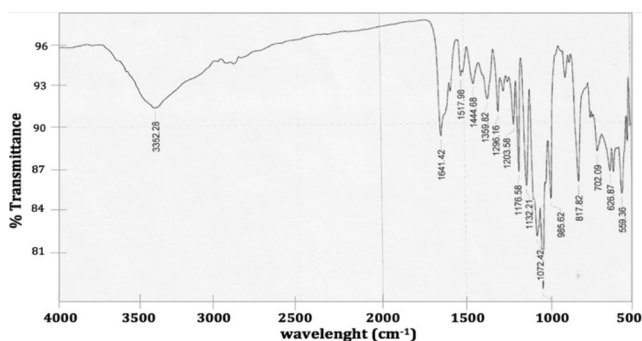


Table 1 shows the values obtained from statistical analysis of antioxidant references and naringin ( $5.0 \times 10^3$  ppm). Results indicate naringin has lower scavenging free radical activity that reference antioxidant references, however, values are very close, this is an important result due to *Citrus aurantium* peel (source of naringin) is an inexpensive and abundant resource, results indicate that *Citrus aurantium* peel has high potential practical application to orange peel waste. The antioxidant capacity of extracts of citrus fruits is due to the presence of phenolic acids, flavonoids and other phenolic compounds, as generally reported in the literature [18-20]. The results for antioxidant activity of naringin against hydroxyl radicals by ABTS<sup>•+</sup> verified that *Citrus aurantium* peel has potential antioxidant activity against free radical; this an important result due to *Citrus aurantium* peel is cheap and abundant natural resource in our Caribbean region.

### 4. CONCLUSIONS

**Table 1.** Scavenging free radicals of naringin and references patterns.

**Tabla 1.** Actividad antioxidante contra radicales libres del extracto de Naringina y los patrones de comparación.

Sample	Absorbance*	Standard Deviation	CV**	SFR (%)
TROLOX 3.5x10 <sup>2</sup> ppm	0,33	0,02	5	50,4
BHT 3.5x10 <sup>2</sup> ppm	0,34	0,02	6	49,4
VIT E 5.5x10 <sup>2</sup> ppm	0,31	0,02	8	52,9
Narigin 5x10 <sup>3</sup> ppm	0,312	0,007	2	43,1

\*After 30 min

\*\*Variation Coefficient, %

In this work we isolated naringin from *Citrus aurantium*, the analysis through UV-vis and IR-FT confirmed the identity of compound. Furthermore, we showed that naringin is scavenging free radicals. The naringin stabilizes the radical-cation and it had a scavenging free radicals of 43.1 %, this result corresponds to 85.5 % of scavenging free radical of Trolox; Our results indicated that orange peel waste could be used as source of scavenging free radicals.

## 5. ACKNOWLEDGMENTS

W. Vallejo and C. Diaz-Urbe thanks to Universidad del Atlántico for financial support through project of IMPACTO CARIBE 2014 (Resolución 002627 - 3 / Mar/2015).

A. Muñoz thanks to Universidad del Norte for the financial support through of the Strategic Area "Biodiversidad, Servicios Ecosistémicos y Bienestar Humano" (Code Project 2013-DI0024).

## REFERENCES

[1] T. Herraiz, J. Galisteo. "Hydroxyl radical reactions and the radical scavenging activity of B-carboline alkaloids", *Food Chemistry*, 172, 640–649, 2015.

[2] J. B. Jeong, E. W. Seo, H. J. Jeong. "Effect of extracts from pine needle against oxidative DNA damage and apoptosis induced by hydroxyl radical via antioxidant activity", *Food and Chemical Toxicology*, 47, 2135–2141, 2009.

[3] M. Özyürek, B. Bektasoglu, K. Güclü, R. Apak. "Hydroxyl radical scavenging assay of phenolics and flavonoids with a modified cupric reducing antioxidant capacity (CUPRAC) method using catalase for hydrogen peroxide degradation", *Analytica chimica*

*acta.*, 616, 196–206, 2008.

[4] M. Schrag, C. Mueller, M. Zabel, A. Crofton, W.M. Kirsch, O. Ghribi, R. Squitti, G. Perry. "Oxidative stress in blood in Alzheimer's disease and mild cognitive impairment: A meta-analysis", *Neurobiology of Disease*, 59, 100-110, 2013.

[5] G. Chen, L. Ren, J. Zhang, B. M. Reed, D. Zhang, X. Shen. "Cryopreservation affects ROS-induced oxidative stress and antioxidant response in Arabidopsis seedlings", *Cryobiology*, 70, 38–47, 2015.

[6] P. Sunkireddy, S. N. Jhab, J. R. Kanwar, S. C. Yadav. "Natural antioxidant biomolecules promises future nanomedicine based therapy for cataract", *Colloids and Surfaces B: Biointerfaces*, 112, 554–562, 2013.

[7] F. M.F. Roleira, E. J. Tavares, C. L. Varela, S. C. Costa, T. Silva, J. Garrido, F. Borges. "Plant derived and dietary phenolic antioxidants: Anticancer properties", *Food Chemistry*, 183, 235–258, 2015.

[8] J. Li, S. Fan, Z. Qiu, C. Li, S. Nie. "Total flavonoids content, antioxidant and antimicrobial activities of extracts from Mosla chinensis Maxim. cv. Jiangxiangru", *LWT - Food Science and Technology*, 64, 1022-1027, 2015.

[9] M. Iranshahi, R. Rezaee, H. Parhiz, A. Roohbakhsh, F. Soltani. "Protective effects of flavonoids against microbes and toxins: The cases of hesperidin and hesperetin", *Life Sciences*, 137, 125–132, 2015.

[10] D. Ravishankar, A. K. Rajora, F. Greco, Helen. M.I. Osborn. "Flavonoids as prospective compounds for anti-cancer therapy", *The International Journal of Biochemistry & Cell Biology*, 45, 2821–2831, 2013.

[11] M. Bacanl, A. A. Başaran, N. Başaran. "The

antioxidant and antigenotoxic properties of citrus phenolics limonene and naringin", *Food and Chemical Toxicology*, 8, 160–170, 2015.

[12] T. H. Kim, S. J. Jang, H. Chung, H. Kim, H. Yong, W. Choe, C. Jo. "Enhancement of antioxidant effects of naringin after atmospheric pressure dielectric barrier discharge plasma treatment", *Bioorganic & Medicinal Chemistry Letters*, 25, 1236–1239, 2015.

[13] P. Thangavel, R. Muthu, M. Vaiyapuri, "Antioxidant potential of naringin – a dietary flavonoid – in N-Nitrosodiethylamine induced rat liver carcinogenesis". *Biomedicine & Preventive Nutrition*, 2, 193–202, 2012.

[14] Ikan R. Acetogenins in Natural Products, London and New York editors, Academic Press, p. 45-60.

[15] N. Miller, C. Rice-Evans. "Factors Influencing the Antioxidant Activity Determined by the ABTS<sup>•+</sup> Radical Cation Assay", *Free Radical Research*, 26, 195-9, 1997.

[16] Liangli L. Yu (Editor). *Wheat Antioxidant*. New

York, Wiley editors, p. 129.

[17] M. Özyürek, D. Akpınar, M. Bener, B. Türkan, K. Güçlü, & R. Apak, "Novel oxime based flavanone, naringin-oxime: Synthesis, characterization and screening for antioxidant activity". *Chemico-Biological Interactions*, 212, 40-46, 2014.

[18] P.L. Muthiah, M. Umamaheswari, K. Asokkumar "In vitro antioxidant activities of leaves, fruits and peel extracts of Citrus", *International Journal of Phytopharmacy*, 2, 13-20, 2012.

[19] V. Roginsky, E.A. Lissi, "Review of methods to determine chain-breaking antioxidant activity in food". *Food Chemistry*, 92, 235-254, 2005.

[20] A. Bocco, M. E. Cuvelier, H. Richard, C. Berset. "Antioxidant activity and phenolic composition of citrus peel and seed extracts", *Journal of Agricultural Food Chemistry*, 46, 2123-2129, 1998.